

A study on cross-talk nerve stimulation: electrode placement and current leakage lid

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Abstract - Cross-talk phenomena should be avoided when stimulating nerves. One option to limit the current spread is to use tripolar electrodes, but at the cost of increasing the number of wires connection. We investigate the impact of the central electrode position and of current spread on cross-talk, in a set of tripolar or quasi-tripolar configurations.

I. INTRODUCTION

Tripolar electrode arrays are used for nerve stimulation since they limit current spread away from the electrodes, hence allowing better control of the stimulating field.

On the other hand, there is a demand for thin and very compliant cable, even stretchable, to connect electrode arrays to the stimulator, and this is more even true as the number of electrodes needed to achieve functional stimulation will likely increase[1]. This involves reducing the number of wires in the cable, hence decreasing the number of wires per tripole. For one tripole, this can be achieved by connecting the outer electrodes together as a common anode. If several tripoles are used next to one another, all anodes can be connected together as a common anode, further reducing the number of wires from $3n$ to $n+1$, where n is the number of tripole.

When a single voltage source is used, with a common anode configuration, any imbalance in the impedance between the two paths (from either anode to the central cathode) will cause a current imbalance. This can cause activation of neighboring nervous structure, there is cross-talk.

In practice, this was observed in two patients who had been implanted with a LARSI (Lumbar anterior root stimulator implant) [2]. It is important for the design of any new stimulator that we understand the situation and the minimum number of wires to avoid cross-talk stimulation. Hence, we studied this phenomena with a virtual/mathematical model as well as experimental measurements on the sciatic nerve of a *Xenopus Laevis* [3].

II. MATERIAL AND METHOD

A. The LARSI situation

We modeled a LARSI situation to see how our results, experimental and virtual, could reproduce and explain the cross-talk observed with the implanted patient. The LARSI was developed by the Implanted Devices Group of University College of London to restore paraplegic patient's ability to control their lower limb [2]. It is implanted in the cauda equina, where the lumbar roots to be stimulated are available in a single space.

The electrode array used with the LARSI is composed of three tripoles with U-shaped electrodes, in three slots side by side, separated by a thin sheet of silicone rubber, and sealed with

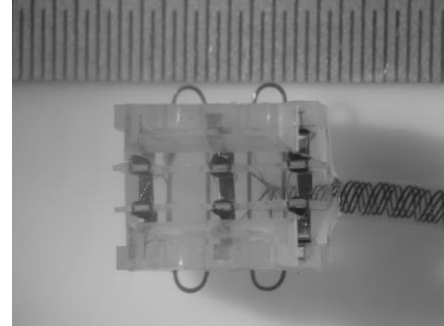


Figure 1. LARSI stimulator with three slot of tripoles [3]

a lid. This array is called a book electrode (Fig. 1). One nerve root was placed in each slot, and the data recorded with 2 patients suggest that stimulation through the electrodes of one slot could cause activation of the nerve root in an adjacent slot. It is this situation that we have modelled here: two nerves in two adjacent slots.

In each slot, the central electrode is used as the cathode while the outer electrodes are anodes. The electrodes in the 3 slots can be connected in 4 different configurations described in Table 1 and sketched on Fig. 2. Situations are (a) Independent between slots - Paired anodes (IP), (b) Common between slots - Paired anodes (CP), (c) Independent between slots - UnPaired anodes (I\P) and (d) Common between slots - UnPaired anodes (C\P).

Table 1. Stimulator configurations for an n -tripoles book

Name	Abbreviation	Number of wires	Figure
Independent between slot - Paired anodes	IP	$2n$	2 (a)
Common between slots - Paired anodes	CP	$n+1$	2 (b)
Independent between slots - UnPaired anodes	I\P	$3n$	2 (c)
Common between slots - UnPaired anodes	C\P	$n+2$	2 (d)

B. Mathematical Model

To evaluate the electrical potential in the nerve root, a finite element model was built in COMSOL Multiphysics.

The equation to be solved is the following, accounting for current conservation,

$$\nabla \cdot \bar{\sigma} \nabla V = 0, \quad (1)$$

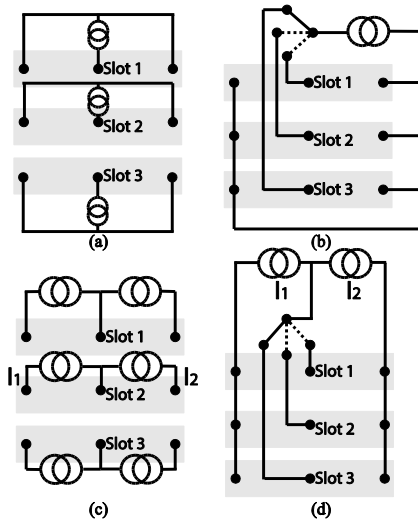


Figure 2. Stimulator configuration for a three slot book - (a) Independent between slots, Paired, anodes (IP); (b) Common between slots, Paired, anodes (CP); (c) Independent between slots, UnPaired, anodes (UP); (d) Common between slots, UnPaired, anodes (UP);

where V is the electrical potential, and $\bar{\sigma}$ the conductivity tensor. Material conductivities are from [4].

This model is coupled to an ODE (ordinary Differential Equation) system modeling the stimulators electrical circuits (Fig. 3).

The activation of nervous fibres inside a root can be evaluated using the Activating Function (AF) [5], i.e.

$$f_n = \frac{d}{4 \rho_i} \frac{V_{n-1} - V_n + V_{n+1} - V_n}{c_m \Delta z L_n}, \quad (2)$$

where d is the diameter of the fibre, ρ_i , the axoplasm resistivity, c_m , the membrane capacitance per unit area, Δz , the discretization length of axon, V_i , the electrical potential at $z = i\Delta z$, and L_n , the internodal distance. The activating function is a valuable tool to check if an action potential is triggered in a nervous fibre at a specific location. Positive peaks of the AF indicate a depolarization of the membrane, that can lead to the creation of an AP. Inversely, negative peaks indicate hyperpolarization. By defining an arbitrary threshold for the activating function, we evaluate which nervous fibre is active, and at which location an action potential will be fired.

The cross-talk ratio (ctr) between nerves is evaluated by computing the ratio between the maximum of the AF along a nervous fibre located on the central line of the nerve being in the slot stimulated and a nerve in an outlying slot (Fig. 3), i.e.

$$ctr = \frac{\max_z(f(z)_{main\ nerve,central})}{\max_z(f(z)_{outlying\ nerve,central})}. \quad (3)$$

A high ctr indicates a lower likelihood of accidental stimulation of neighboring nerves.

The slots of the book electrode are separated by thin sheets of silicone rubber, and the 3 slots are closed by a lid. To evaluate the importance of the lid's seal, and the impact of current spreading between slots, we computed the ctr for different gaps between the walls of the slot and the lid.

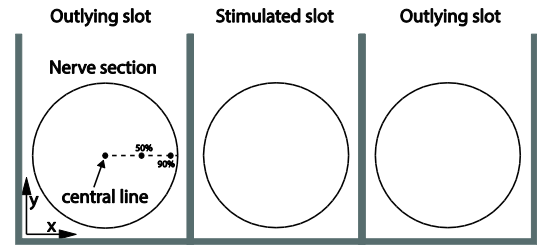


Figure 3. Definition of lines along the nerve where activating function is computed in a nerve section; Highlight of the stimulated/outlying slots.

C. Animal Model

A series of in vitro experiments on the sciatic nerves of *Xenopus Laevis* were performed.

Female frogs were decapitated and pithed prior to dissection. The sciatic nerves on both sides were exposed, from the spinal cord to the knee (common peroneal branch). A thread was then tied at both ends to avoid losing the axoplasm. The threads were also used to handle the nerves as they should be touched as little as possible to preserve their integrity. From the start of the dissection, the nerves were kept moist with amphibian Ringer solution at room temperature. The same care was taken after removal from the body. The explanted nerves were 80 to 100mm long, with a diameter of 1mm at the spinal end and about half that below the knee. Each experiment spanned two days (one day per nerve) as frog nerves carefully handled and kept in Ringer remain excitable for up to 36 hours.

The nerve was mounted in a two-chamber bath as shown in Fig. 4. Using the precision screws, the tension in the nerve was adjusted to hold it just straight (rather than bowing at the middle) without over-stretching it. On the stimulation side, it was placed in the central slot of a book immersed in Ringer solution. On the recording side, it was resting, in air, on a pair of platinum hook electrodes connected to an amplifier (WorldPrecision ISO-DAM8A, gain set to 100 or 1000) and the signal was displayed on an oscilloscope and recorded. The part of the nerve in air was kept moist with Ringer solution, and care was taken to avoid shorting the hooks with the formation of a drop of solution.

The current pulses delivered by the experimental stimulator at a frequency of 1Hz were charge balanced. Their shape was rectangular. Their width was adjustable up to nearly 3ms. The amplitude of the pulse delivered to each anode was controlled. The cathodal current is given by the sum of both anodal currents.

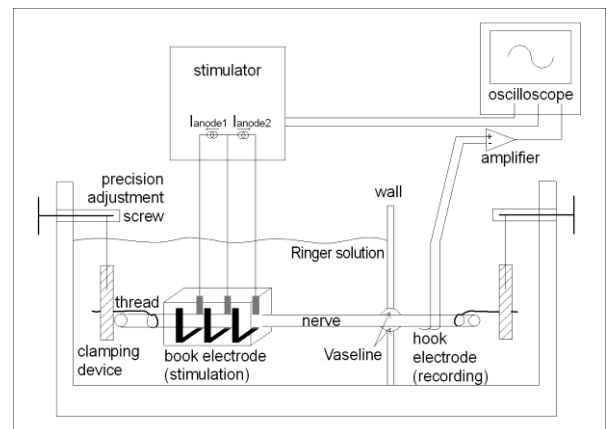


Figure 4. Experimental setup, for clarity only the central slot is shown [3].

The total current amplitude was adjustable with a potentiometer up to 10mA. The current imbalance is calculated as $i_1/(i_1 + i_2)$.

To study the cross-talk threshold, current pulses were sent to the outer slot, and their amplitude increased until compound action potentials from the nerve in the other, passive slot, were recorded at the hook electrodes. Cross-talk ratios were estimated as the ratio of the cross-talk threshold to the direct activation threshold. This was done for all the current imbalances. These tests were repeated on 19 frog nerves.

III. RESULT

A. Mathematical Model

Fig. 5(a) shows AF curves for the CP configuration with perfectly balanced current along three nervous fibres located at the fibre centre, at 50% off-centre, and at 90% off-centre (Fig. 3). The positive peak, indicative of strong depolarization, is at the level of the central cathode. While the two negative peaks are located at the level of the two anodes. Fig. 5(b) shows activating function curves for fibres in a neighboring, non-stimulating, slot. In CP, all external anodes are connected together, hence the hyperpolarization peaks at the level of these electrodes. However, we also observe strong depolarization peaks at the edges of the slot ($z = 0\text{ mm}$ and $z = 9\text{ mm}$). These zones, where action potentials are likely to be elicited, are known as virtual cathodes. Virtual cathodes are present at both ends of the peripheral slot.

B. Animal model

Compound Action Potentials (CAP) were elicited in the nerve in the active slot (see Fig. 6) for all current imbalances. CAP were also elicited in the nerve in the adjacent, passive, slot for the least tripolar of the current imbalances. Average cross-talk ratios computed from these measurement are plotted on Fig. 7.

C. Model validation

Fig. 7 shows the cross-talk ratios computed using the mathematical model for the IP situation (continuous line) and calculated from the sciatic nerve measurement (box plot). The central mark in the box represents the median, the edges are the 25th and 75th percentiles, and the whiskers extend to the most extreme data points not considered outliers. Outliers are represented by crosses.

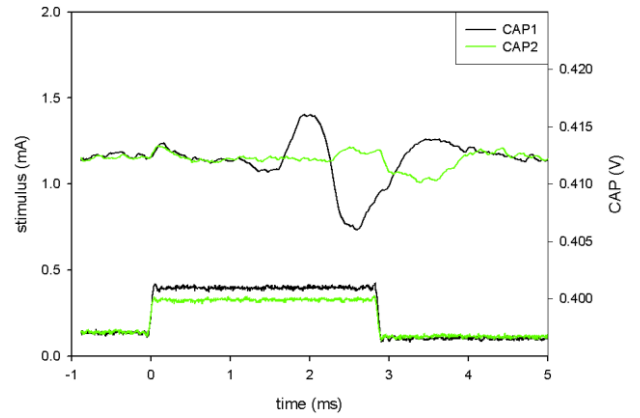
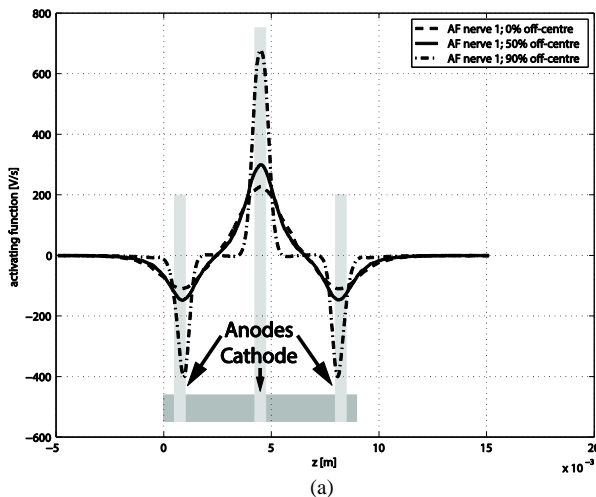


Figure 6. Compound action potential for true tripole (no current imbalance), for a stimulus at threshold (CAP2) and one 16% larger (CAP1).

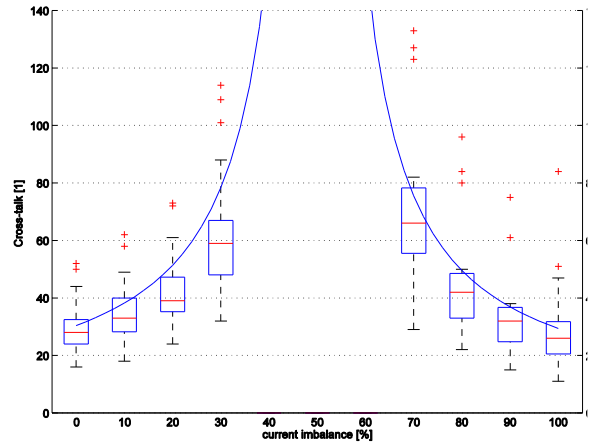


Figure 7. Cross-talk ratio depending of current imbalance for Independent between slots, Unpaired, anodes (IP) - Box plot represent measured data from [3]; continue line represent simulated data.

D. Influence of leaking book

The results presented here are obtained by opening the lid of the book when the current is perfectly balanced to simulate the effect of a leak in the insulation of the electrode mount.

When the lid is perfectly sealed, both configurations have a very different cross-talk ratio (140 for CP and over 3000 for IP). However if a leak is present (lid not well sealed), the cross-talk ratio is radically decreased in both configurations (8 for CP and

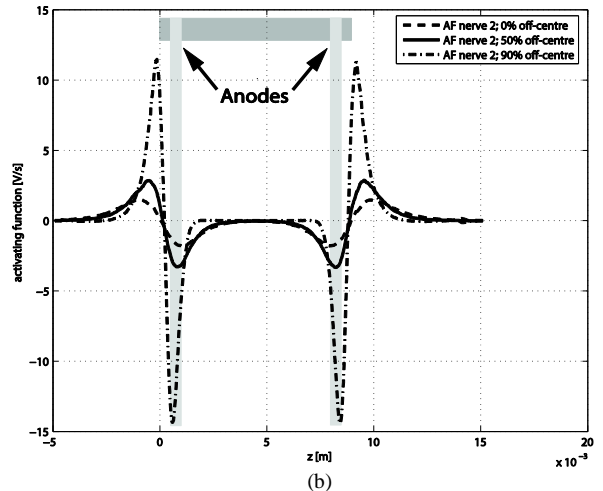


Figure 5. Activating function for nerve fibres at 3 locations inside a nerve. (a) Nerve in active slot. (b) Nerve in peripheral, non-stimulating, slot.

12 for IP) even for a very small leak (gap between the lid and the vertical wall = 63 μm).

E. Influence of central electrode position

The influence of the position of the central electrode on the cross-talk is visible on Fig. 8. The position of the central electrode is described as a relative position in the slot (0% is the location of the first electrode, 50% the normal position, and 100% corresponds to the position of the last electrode).

IV. DISCUSSION

A. Mathematical Model

Figure of the activating function (Fig. 5) show well the direct electrode activation and virtual cathodes present at both end of the slot. This offers a likely explanation for the origin of the cross-talk observed in the patients.

Two facts can be observed on the figures of the activating function:

- The amplitude of the activating function of a nervous fibre in the stimulation slot varies almost three times inside the same nerve depending on the depth of its location.
- There is cross-talk, even if the current is perfectly balanced: virtual electrodes are present on the nerves not being stimulated.

Our results also show that the smallest depolarization peak is always obtained for the fibre along the centre of the nerve. Hence, if this peak is high enough to trigger an action potential, most fibres in the nerves are likely to be activated. However, this will not be the case if there is a hyperpolarization that blocks the action potential occurring near a real or virtual anode.

B. Animal Model

Cross-talk was present, however at ratios larger than to those observed in the LARSI patients [2], and only when the current was not balanced. In none of our 19 experiments did we observe cross-talk when the current imbalance was 10% or less.

C. Model validation

As seen on Fig. 7, the results of the mathematical model are close to those of the animal experiment, validating our mathematical model as an acceptable representation of an *in vitro* situation. In the future, this mathematical model will be used to

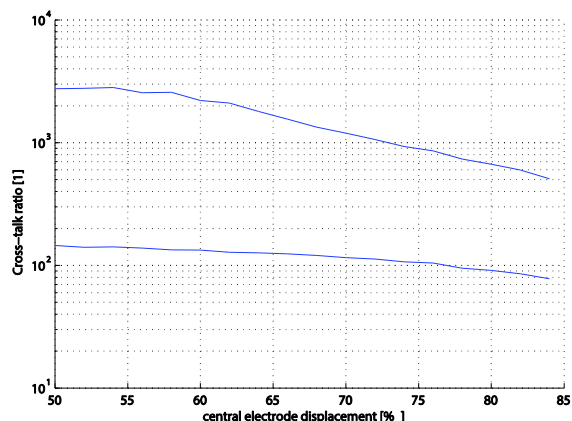


Figure 8. Comparison of cross-talk ratio for electrode displacement for IP (upper curve) and CP (lower curve).

model more electrode configurations, causes of current imbalance, and other parameters not easily controlled in an experimental setting.

D. Influence of leaking book

The results show that care should be taken when sealing the slot as a leak can considerably decrease the cross-talk threshold. This suggests that in the LARSI, where the cross-talk ratio was below 10, there might have been some current spread caused by poor insulation, maybe combined with an imbalance of the current.

A possible reason for this would be an unequal growth of connective tissue, that could have affected the lead seal (as it was not glued in place), and caused the impedance between the electrodes to be considerably different.

E. Influence of the central electrode position

We have only begun to study the influence of central electrode displacement. A misplacement of the central electrode relative position up to 20% is quite likely for hand-built commercial electrodes. Fig. 8 shows that this would have reduced by three the cross-talk ratio of independent slot with paired anode, but in regards of the high cross-talk ratio for current balanced situation (relative position of 50%), this is not an issue.

V. CONCLUSION

Reducing the number of wires from $2n$ to $n+1$ (IP to CP configuration) increases the cross-talk even if the current is perfectly balanced. However, the cross-talk ratio when the current is perfectly balanced is very high in both configurations. If the current is not balanced, and is due to central electrode misplacement, the decrease of *ctr* in both configuration is still acceptable. A central electrode misplacement of a few percent is almost negligible for *ctr*. A leak in the seal, even with balanced current, is much worse with regards to the cross-talk.

In conclusion, using a CP configuration ($n+1$ wires) is acceptable for *ctr* if the lid is perfectly sealed.

Further investigation should focus on other parameters: impedance asymmetry due to contamination of the slot by a material of different conductivity, imperfect geometry of the book, etc.

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REFERENCES

- [1] P. R. Troyk and N. de N. Donaldson, "Implantable FES Stimulation Systems: What is Needed?," *Neuromodulation*, vol. 4, no. 4, pp. 196–204, Oct. 2001.
- [2] N. Donaldson, D. Rushton, and T. Tromans, "Neuroprostheses for leg function after spinal-cord injury," *Lancet*, 1997.
- [3] A. Vanhoestenbergh, "Implanted Devices : Improved Methods for Nerve Root Stimulation," University College of London, 2007.
- [4] L. A. Geddes and L. E. Baker, "The specific resistance of biological material—A compendium of data for the biomedical engineer and physiologist," *Med. Biol. Eng.*, vol. 5, no. 3, pp. 271–293, May 1967.
- [5] F. Rattay, "Analysis of Models for Extracellular Fiber Stimulation," *IEEE Trans. Biomed. Eng.*, vol. 36, no. 7, pp. 676–682, 1989.